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Agaricus globocystidiatus: a new neotropical species with pleurocystidia in *Agaricus* subg. *Minoriopsis*

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Abstract

Agaricus is a monophyletic genus with a worldwide distribution and more than 400 described species. The genus grows on soil and can be easily recognized by the presence of an annulus on the stipe and free lamellae which become dark brown with spore maturation. Although *Agaricus* is easily recognized in the field because of its macroscopic characters, identification at the species level is difficult. Based on specimens collected in the states of Paraná and Santa Catarina, in the south of Brazil, we propose a new species *Agaricus globocystidiatus*. The new taxon is distinguished mainly by the presence of pleurocystidia, a rare morphological character in *Agaricus*. Molecular analyses based on nuc rITS1-5.8-ITS2 (ITS) barcode sequences show that *A. globocystidiatus* belongs to *Agaricus* subg. *Minoriopsis*.

Keywords: Agaricaceae, Brazil, phylogeny, systematics

Introduction

The genus Agaricus L. (1753:1171) (Basidiomycota) is monophyletic (Vellinga 2004) and comprises approximately 400 species worldwide (Karunarathna et al. 2016). Linnaeus (1753) was the first to describe the genus, in a very broad sense, including all mushrooms with lamellae and a stipe. Due to this, many species of mushroom-forming fungi were first described as Agaricus and have this genus name as their basionym. The representatives of Agaricus are saprobic and usually have the following features: large and fleshy basidioma, with a white, yellow, brown or sometimes blackish or purple pileus, which is easily separable from the stipe; lamellae free, white when young, changing to pink and finally becoming brown to dark brown when the basidiospores are mature; and an annulus that is always present and usually membranous, white, and sometimes double (Heinemann 1956; Singer 1986; Zhao et al. 2016). Macroscopic characters for the genus can be easily recognized in the field but species delimitation is very complex. Environmental factors, intraspecific variability and lack of characters to differentiate species are some of the problems found in the taxonomy of this genus (Zhao et al. 2011). Agaricus is microscopically characterized by the pileipellis being a cutis, hymenophoral trama regular to slightly interwoven, hyphae without clamp connections, brown basidiospores without a true apical pore, often the absence of pleurocystidia and presence or absence of cheilocystidia (Heinemann 1956; Singer 1986; Zhao et al. 2016). In addition to macroscopic and microscopic features, the Schäffer reaction and organoleptic characters (e.g. odor) are useful to identify subgenera, sections and species (Heinemann 1956; Zhao et al. 2016; Chen et al. 2017).

Molecular data and improved phylogenetic studies have helped clarify relationships among *Agaricus* taxa since Mitchell & Bresinsky (1999) produced the first phylogenetic analysis of the genus using ITS sequence data, based on ITS2 and 28S rDNA. Zhao *et al.* (2011) were the first to study molecular data from tropical specimens and observed that only about one-third of tropical species belong to the classical sections historically proposed based on

temperate species, which suggests that the systematics of the genus need to be expanded. Zhao *et al.* (2016) analyzed morphological and molecular data from more than 700 samples and proposed a reconstruction of *Agaricus* based on divergence time to define taxonomic ranks. They produced a multi-gene phylogeny combining ITS, LSU, *tef-1a* and *rpb2* sequences of samples from tropical and temperate areas, and segregated the genus into five subgenera and twenty sections. Zhao *et al.* (2016) included in *Agaricus* subg. *Minores* (Fr.) R.L. Zhao & Moncalvo *Agaricus* sect. *Minores* (Fr.) Henn. and *Agaricus* sect. *Laeticolores* Heinem.

Recently, Chen *et al.* (2017) studied *Agaricus* subg. *Minores* and allied clades. According to Chen *et al.* (2017) the clade representing *Agaricus* sect. *Laeticolores* Heinem. in Zhao *et al.* (2016) corresponds to a new subgenus because *Agaricus* sect. *Laeticolores* is in fact a synonym of *Agaricus* sect. *Minores* (Fr.) Henn. since the type species of *Agaricus* sect. *Laeticolores*, *Agaricus* laeticulus Callac, L.A. Parra, Linda J. Chen & Raspé (2017:182), belongs to *Agaricus* sect. *Minores*. In agreement with the taxonomic system based on divergence time (Zhao *et al.* 2016) Chen *et al.* (2017) proposed *Agaricus* subg. *Minoriopsis* Linda J. Chen, L.A. Parra, Callac, Angelini & Raspé and ranked as subgenus because it has a stem age of 31.02 Ma. *Agaricus* subg. *Minoriopsis* is more related to *Agaricus* subg. *Flavoagaricus* Subg. *Minores* (Fr.) R.L. Zhao & Moncalvo. *Agaricus* subg. *Minores* includes *Agaricus* subg. *Minores* (Fr.) Henn, *Agaricus* sect. *Leucocarpi* Linda J. Chen & Callac, and one unnamed section 1 (Chen *et al.* 2017).

The specimens of *Agaricus* subg. *Minoriopsis* are characterized by having a dark reddish purple, rarely reddish brown Schäffer's reaction; a positive KOH reaction; an odor of anise or almonds; a superous annulus that is thick at the margin, double, fibrillose-squamose, or sometimes with squamules radially arranged on the lower surface; and cheilocystidia that are clavate, pyriform, more or less globose or fusiform (Chen *et al.* 2017).

The main studies that describe species of *Agaricus* from Brazil were published by Pegler (1990), Heinemann (1993) and Meijer (2008). Pegler (1990) revised some specimens of agaricoid fungi from Brazil, collected by J.P.F.C. Montagne in 1843, and reported three species of *Agaricus* with holotypes deposited in the herbarium of the Muséum National d'Histoire Naturelle in France. Heinemann (1993) studied the tribe Agariceae in Brazil and described a new species, *A. meijeri* Heinem. (1993:368), collected in the state of Paraná, and Meijer (2008) described another *Agaricus* from Paraná, *A. stijvei* de Meijer (2008:302). Unfortunately, there are no available sequences from these specimens. To better understand the species distribution and infrageneric relationships in the genus it is important to collect and review specimens from neotropical regions that have been under studied (Zhao *et al.* 2011). In this paper a new species of *Agaricus* subg. *Minoriopsis* is described based on morphological and molecular data.

Materials and methods

Sampling and morphological study

The specimens studied were collected in the states of Paraná and Santa Catarina, in the south of Brazil, and were described macroscopically following Largent (1986) and Vellinga (1988). The chemical reactions were performed with 3% KOH (tissue turns yellow when positive) and 10% NaOH (tissue turns orange-red when positive) on the pileus and stipe surface of fresh or dried basidiomata. The color code is based on the *Online Auction Color Chart* (Kramer 2004). Micromorphological features were observed from dried specimens following the nomenclature terms of Largent & Thiers (1977) and Vellinga (1988). The notation [a/b/c] at the beginning of a set of basidiospores represents the following: (a) basidiospores were measured from (b) basidiomata taken from (c) collections. Basidiospore measurements were based on at least 20 structures per basidioma, including the Q-value (quotient of length to width), Q_m (the mean of the Q-value), L_m (the mean of length), and W_m (the mean of width). Basidiospore shape was based on the Q-value (Bas 1969).

DNA extraction, amplification and sequencing

DNA was extracted from dried pileus and lamellae fragments, following a modified CTAB extraction method (Góes-Neto *et al.* 2005). The nuc rITS1-5.8S-ITS2 (ITS) sequences were generated using the primer pair ITS8F and ITS6R (Dentinger *et al.* 2010). The PCR mix was prepared using4 µL genomic DNA (1:10), 10 µL water, 10 µL Master mix (Promega) and 0.5 µL each primer (10pmol/L). The PCR program was according to Zhao *et al.* (2011): 5 min at 95 °C; 35 cycles (1 min. at 94 °C, 1.5 min. at 55 °C, 1.5 min. at 72 °C); and 5 min at 72 °C. The PCR products were purified with 20% PEG and sequenced at Centro de Pesquisas René Rachou (Fiocruz Minas, Brazil) using the same primer pair.

Phylogenetic analyses

The generated sequences were manually checked and edited with Geneious v.8.1 (Kearse et al. 2012). Our dataset comprises sequences from eight newly collected Agaricus specimens from Brazil, 29 sequences of Agaricus subg. Minores and ten sequences of Agaricus subg. Minoriopsis taxa from GenBank (Table 1). The sequence of A. albosquamosus L.J. Chen, K.D. Hyde & R.L. Zhao (2016:46) (LD2012192), which is part of Agaricus subg. Spissicaules (Heinem.) R.L. Zhao & Moncalvo, was used as the outgroup. Newly obtained sequences were deposited in GenBank and accession numbers and country of origin are provided in Table 1. Sequences were aligned in MAFFT v7 (Katoh 2013) following the Q-INS-I criteria and then manually corrected using MEGA v7 (Tamura et al. 2013). To identify the best nucleotide evolution model, we used the AIC criterion (Akaike Information Criterion) in jModelTest v2.1.6 (Guindon et al. 2010; Darriba et al. 2012). The ITS region was partitioned into ITS1, 5.8S and ITS2, and the evolution model was estimated for each partition. The maximum likelihood (ML) analysis was performed in RAxML v8.2.8 (Stamatakis 2014) using the GTRGAMMA model with a rapid bootstrap analysis with 1,000 replicates and search for the best-scoring ML tree, and other parameters estimated by the software. The Bayesian inference (BI) analysis was performed with Mr.Bayes v3.2.6 (Ronquist & Huelsenbeck 2003) using three partitions (ITS1, 5.8S, ITS2), with two independent runs, four simultaneous independent chains and 20,000,000 generations with a sample frequency every 1,000 generation. RAxML, jModelTest, and Mr.Bayes were used in the CIPRES Science Gateway 3.1 (Miller et al. 2010). The outputs of the analyses were displayed with FigTree v1.4.2 (http://tree.bio.ed.ac.uk/software/figtree/).

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Agaricus sp.ZRLWXH3150KT951390China, GuangdongZhao et al. (2016)Agaricus comtulusLAPAG724KT951332Spain, BurgosZhao et al. (2016)Agaricus gemliiAH-44510KF47891Spain, CanariasParra (2013)Agaricus pallensLAPAG926KT951315Sweden, LaplandZhao et al. (2016)Agaricus sp.CA846JF727865ThailandZhao et al. (2016)Agaricus sp.ZRL2012357KT951369China, YunnanZhao et al. (2016)Agaricus sp.ZRL2012576KT951372China, JiangxiZhao et al. (2016)Agaricus sp.ZRL2012714KT951381China, TibetZhao et al. (2016)Agaricus sp.ZRL2012714KT951381China, TibetZhao et al. (2016)Agaricus sp.ZRL201039KT951351China, TibetZhao et al. (2016)Agaricus sp.ZRL2011039KT951351China, TunnanZhao et al. (2016)Agaricus sp.ZRL2012714KT951329Spain, CastellónZhao et al. (2016)Agaricus sp.ZRL2012012KT951351China, YunnanZhao et al. (2016)Agaricus sp.ZRL2012012KT951359China, YunnanZhao et al. (2016)Agaricus sp.ZRL2012199KT951367China, YunnanZhao et al. (2016)Agaricus sp.ZRL2012199KT951391China, GuangdongZhao et al. (2016)Agaricus sp.ZRL2012129KT951391China, GuangdongZhao et al. (2016)Agaricus sp.ZRL2012129KT951391China, Guangdong	Agaricus sp.	ZRLWXH3076	KT951388	China, Fujian	Zhao et al. (2016)
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Agaricus sp.ZRL2012576KT951372China, TibetZhao et al. (2016)Agaricus sp.ZRL2012714KT951381China, TibetZhao et al. (2016)Agaricus sp.ZRL2011039KT951351China, YunnanZhao et al. (2016)Agaricus aridicolaLAPAG589KT951331Spain, CastellónZhao et al. (2016)Agaricus pseudolutosusLAPAG454KT951329Spain, BurgosZhao et al. (2016)Agaricus sp.ZRL2012012KT951359China, YunnanZhao et al. (2016)Agaricus sp.ZRL2012012KT951367China, YunnanZhao et al. (2016)Agaricus sp.ZRL2012199KT951367China, YunnanZhao et al. (2016)Agaricus sp.ZRLWXH3161KT951391China, GuangdongZhao et al. (2016)Agaricus sp.ZRLWXH3161KT951312Dominican Republic, SosúaZhao et al. (2016)Agaricus sp.LAPAM14KT951312Dominican Republic, SosúaZhao et al. (2016)Agaricus sp.LAPAM14KT951355ThailandZhao et al. (2016)Agaricus subg. MinoriopsisMATA816JF727870Mexico, VeracruzZhao et al. (2011)Agaricus globocystidiatusMPD02MF188248Brazil, Santa CatarinaAgaricus globocystidiatusMPD04Agaricus globocystidiatusMPD09MF188247Brazil, Santa CatarinaAgaricus globocystidiatusMPD128Agaricus globocystidiatusMPD18MF188249Brazil, Santa CatarinaAgaricus globocystidiatusMPD128	Agaricus sp.	ZRLWXH3067	KT951387	China, Jiangxi	Zhao et al. (2016)
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Agaricus sp.ZRL2011039KT951351China, YunnanZhao et al. (2016)Agaricus aridicolaLAPAG589KT951331Spain, CastellónZhao et al. (2016)Agaricus pseudolutosusLAPAG454KT951329Spain, BurgosZhao et al. (2016)Agaricus sp.ZRL2012012KT951359China, YunnanZhao et al. (2016)Agaricus sp.ZRL2012199KT951367China, YunnanZhao et al. (2016)Agaricus sp.ZRL2012199KT951367China, YunnanZhao et al. (2016)Agaricus sp.ZRLWXH3161KT951391China, GuangdongZhao et al. (2016)Agaricus sp.ZRLWXH3161KT951312Dominican Republic, SosúaZhao et al. (2016)Agaricus sp.LAPAM14KT951355ThailandZhao et al. (2016)Agaricus candidolutescensLD2012129KT951355ThailandZhao et al. (2016)Agaricus globocystidiatusMATA816JF727870Mexico, VeracruzZhao et al. (2011)Agaricus globocystidiatusMPD03MF188248Brazil, Santa CatarinaZhao et al. (2011)Agaricus globocystidiatusMPD04MF188247Brazil, Santa CatarinaJerzil, Santa CatarinaAgaricus globocystidiatusMPD29MF188249Brazil, Santa CatarinaJerzil, Santa Catarina	Agaricus sp.	ZRL2012714	KT951381	China, Tibet	Zhao et al. (2016)
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Agaricus sp.ZRL2012199KT951367China, YunnanZhao et al. (2016)Agaricus callaciiAH-42929KF447899Spain, CanariasParra (2013)Agaricus sp.ZRLWXH3161KT951391China, GuangdongZhao et al. (2016)Agaricus sp.LAPAM14KT951312Dominican Republic, SosúaZhao et al. (2016)Agaricus candidolutescensLD2012129KT951355ThailandZhao et al. (2016)Agaricus subg. Minoriopsis </td <td>Agaricus sp.</td> <td>ZRL2012012</td> <td>KT951359</td> <td>China, Yunnan</td> <td>Zhao et al. (2016)</td>	Agaricus sp.	ZRL2012012	KT951359	China, Yunnan	Zhao et al. (2016)
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Agaricus candidolutescensLD2012129KT951335ThailandZhao et al. (2016)Agaricus subg. Minoriopsis </td <td>Agaricus sp.</td> <td>LAPAM14</td> <td>KT951312</td> <td>Dominican Republic, Sosúa</td> <td>Zhao et al. (2016)</td>	Agaricus sp.	LAPAM14	KT951312	Dominican Republic, Sosúa	Zhao et al. (2016)
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Agaricus globocystidiatusMPD03MF188248Brazil, Santa CatarinaAgaricus globocystidiatusMPD04MF188245Brazil, Santa CatarinaAgaricus globocystidiatusMPD29MF188247Brazil, Santa CatarinaAgaricus globocystidiatusMPD128MF188249Brazil, Santa Catarina	Agaricus globocystidiatus	MPD02	MF188244	Brazil, Santa Catarina	
Agaricus globocystidiatusMPD04MF188245Brazil, Santa CatarinaAgaricus globocystidiatusMPD29MF188247Brazil, Santa CatarinaAgaricus globocystidiatusMPD128MF188249Brazil, Santa Catarina	Agaricus globocystidiatus	MPD03	MF188248	Brazil, Santa Catarina	
Agaricus globocystidiatusMPD29MF188247Brazil, Santa CatarinaAgaricus globocystidiatusMPD128MF188249Brazil, Santa Catarina	Agaricus globocystidiatus	MPD04	MF188245	Brazil, Santa Catarina	
Agaricus globocystidiatusMPD128MF188249Brazil, Santa Catarina	Agaricus globocystidiatus	MPD29	MF188247	Brazil, Santa Catarina	
	Agaricus globocystidiatus	MPD128	MF188249	Brazil, Santa Catarina	

TABLE 1. Specimens and sequences used for the molecular analyses. Sequences in **bold** were generated during this work.

TABLE 1. (Continued)

Taxon	Collection	ITS	Location	Reference
Agaricus globocystidiatus	MPD19	MF188253	Brazil, Santa Catarina	
Agaricus globocystidiatus	MPD71	MF188251	Brazil, Paraná	
Agaricus globocystidiatus	EC31	MF188252	Brazil, Santa Catarina	
Agaricus sp.	F1779	JF727853	France, Martinique	Zhao et al. (2011)
Agaricus sp.	RMC-1257	KM349612	USA, Arizona	Kerrigan (2016)
Agaricus sp.	RMC-1256	KM349611	USA, Arizona	Kerrigan (2016)
Agaricus rufoaurantiacus	LAPAM15	KT951313	Dominican Republic, Sosúa	Zhao et al. (2016)
Agaricus aff. rufoaurantiacus	CL/GUAD05.099	JF727857	France, Guadeloupe	Zhao et al. (2011)
Agaricus sp.	HAI0386	AJ884624	USA	Didukh et al. (2005)
Agaricus martinicensis	F2815	JF727855	France, Martinique	Zhao et al. (2011)
Agaricus martinicensis	LAPAM16	KX671699	Dominican Republic	Chen et al. (2017)
Agaricus sp.	LAPAM28	KX671700	Dominican Republic	Chen et al. (2017)
Agaricus subg. Spissicaules				
Agaricus albosquamosus	LD2012192	KT951394	Thailand	Zhao et al. (2016)

Results and Discussion

Phylogeny

The final ITS alignment had a total of 689 characters (gaps included) with 47 ingroup sequences, including eight generated in this study. The best models of nucleotide substitution estimated for each partition in the datasets were: TVM+G to ITS1, JC to ITS 5.8S, and HKY+I+G to ITS2. Both ML and BI tree topologies were the same, with differences only among sequences within *Agaricus* subg. *Minores*. The BI obtained by the analysis is presented in Figure 1, with posterior probability (PP) and bootstrap (BS) values above the branches.



FIGURE 1. Bayesian inference tree of *Agaricus* subg. *Minoriopsis* and *Agaricus* subg. *Minores* based on ITS data and rooted with *Agaricus albosquamosus*. The sequences generated in this work are in **bold**. Posterior probability (PP) (above 0.7) and bootstrap (BS) (above 50%) support values are shown above the branches (PP/BS).

Sequences of the new species clustered in a well-supported clade with one sequence from Mexico (JF727870), which probably also represents *A. globocystidiatus*, and other sequences of *Agaricus* subg. *Minoriopsis*. The sequence JF727870 from Mexican material (MATA816) clustered in the clade TRII of Zhao *et al.* (2011) with sequences of five other specimens, but the authors did not assign a taxonomic name to MATA816 and it was not included in further analyses by Zhao *et al.* (2016).

Our analysis focused on the phylogenetic relationships between the new species and closely related species. All known members of *Agaricus* subg. *Minoriopsis* were included and the analysis clearly indicated that the new species belongs to this subgenus. However, in our tree, phylogenetic relationships between taxa of higher rank are not well supported. In our analysis, *Agaricus* subg. *Minoriopsis* appears as a sister group to *Agaricus* sect. *Minores*, and *Agaricus* subg. *Minoriopsis* to what Chen *et al.* (2017) found in a multi-gene phylogenetic analysis. Chen *et al.* (2017) reported eight species in *Agaricus* subg. *Minoriopsis* and we add *A. globocystidiatus* in the present study. All species known to *Agaricus* subg. *Minoriopsis* are from the Americas (Chen *et al.* 2017).

Taxonomy

Agaricus globocystidiatus Drewinski & M.A.Neves *sp. nov.* (Figs. 2, 3) Mycobank: MB821296

FIGURE 2. Microcharacters of *Agaricus globocystidiatus* FLOR 61594 (*MPD29*–Holotype). a. Pileipellis; b. Basidia; c. Cheilocystidia; d. Pleurocystidia; e. Basidiospores. Scale bars: 5µm. Drawings by M.P.Drewinski.

Diagnosis:—Similar to *Agaricus pleurocystidiatus* by the presence of pleurocystidia, but differing in the pileus surface covered by concentrically arranged purple scales, regular pileus margin, and stipe with brown scales and slightly yellowish on some parts when exposed.

Etymology:—*glob*-(L.) refers to the main shape of the pleurocystidia and *cystidium* (L.) refers to the presence of pleurocystidia, a rare morphological character in this genus.

Holotype:—BRAZIL. Santa Catarina: Florianópolis, Universidade Federal de Santa Catarina, near to the Department of Botany, 10 December 2015, *M.P. Drewinski MPD29* (FLOR61594).

Description:—*Pileus* 30–122 mm diam., at first parabolic, then conical truncated to convex and finally expanding to plano-convex, slightly depressed, covered by concentrically arranged purple (oac527) scales on beige (oac780) background, darker at center with concentrated scales that are scattered towards the margin, margin regular. *Pileus context* approx. 5mm thick at center and 1mm at margin, whitish or slightly yellowish in some parts when exposed, fleshy. *Lamellae* free, white at first, then pinkish and finally brown when mature, margin regular, crowded, with lamellulae. *Stipe* 102–120 × 9–20 mm, central, tapering at base, fistulose, beige pearly (oac767), smooth above annulus, with brown scales below, changing to yellow in some parts including the context of the base. *Annulus* superous, pendant, membranous, floccose on lower surface, whitish.

Basidiospores [200/10/8] (4.0–)5.0–6.2(–7.5) × (3.7–)5.0 μ m, [Q=1.25–1.66; Q_m=1.42; L_m=5.3 μ m; W_m=4.2 μ m], broadly ellipsoid to ellipsoid, dark brown, smooth, thick-walled, apiculus conspicuous, without a germ pore. *Basidia* (16.2–)21–37(–40) × 6.2–10.0(–11.2) μ m, narrowly utriform to clavate, hyaline, smooth, 4-spored. *Pleurocystidia* (25–)26–43 × (16.2–)18.7–23 μ m, globose, subglobose, ovoid to broadly clavate, hyaline, smooth, thin-walled. *Cheilocystidia* (11.2–)13.7–45(–47) × (7.5–)10–21(–23) μ m, clavate, subglobose to obovoid, hyaline, thin-walled, usually covering all the lamellae edge. *Lamellar trama* regular, with abundant oleiferous hyphae, up to 6.5(–8.7) μ m diam. *Pileipellis* a cutis composed of cylindrical and thin- to rather thick-walled hyphae, (3.7–)5.0–11.2 μ m diam., branched, with purple internal plasmatic pigments. *Pileus context* composed of cylindrical and thin-walled hyphae, 5.0–10.0 μ m diam., sometimes inflated. *Macrochemical reactions* yellow in 3% KOH (positive), yellow in 10% NaOH (positive). Schaffer's reaction was unclear, but the pileus turned purple with citric acid. *Odor* sweet almond.

Habit, habitat and distribution:—solitary to gregarious, terrestrial, growing on grass in urban areas and areas of the Atlantic Forest domain; Brazil, Santa Catarina and Paraná states.

Specimens examined:—BRAZIL. Santa Catarina: Florianópolis, at the University Campus near the Botany Department, 30 July 2015, *M.P. Drewinski MPD02* (FLOR61586–Paratype); 05 October 2015, *M.P. Drewinski MPD03* (SP466715–Paratype); 07 October 2015, *M.P. Drewinski MPD04*;16 March 2016, *M.P. Drewinski MPD128* (FLOR61612–Paratype); 09 February 2017, *M.P. Drewinski MPD145* (FLOR61620–Paratype); Parque Ecológico do Córrego Grande, 11 February 2016, *E.Copini EC31* (FLOR61583); Cacupé, *Tim Brightwell MPD19* (FLOR61582); Paraná: Guarapuava, Parque Municipal das Araucárias, 12 January 2016, *M.P. Drewinski MPD71*(FLOR61599).

Notes:—The presence of pleurocystidia is a rare morphological character in *Agaricus*. Although those pleurocystidia are large and stand out from the other structures in the hymenium, they are not abundant and may go unnoticed if the material is not carefully studied (Heinemann 1980). *Agaricus globocystidiatus* is similar to *A. pleurocystidiatus* Heinem. (1980:12), from Singapore, mainly due to the presence of pleurocystidia and because of the similar annulus that is described as membranous, pendant, white and with flakes on the lower side (Heinemann 1980). However, *A. pleurocystidiatus* differs by the following: the pileus surface, which has dark brown scales on a light brown background; the pileus margin, which is appendiculate; and the fibrillose light brown stipe. Furthermore, *A. pleurocystidiatus* changes to reddish brown when cut and is known only from Singapore (Heinemann 1980).

Agaricus sinodeliciosus Z.R. Wang & R.L. Zhao (2015:192) from China also has pleurocystidia (Wang *et al.* 2015). However, *A. sinodeliciosus* differs by its semi-hypogeous habit (growing buried in sandy soil), pileus surface covered with light brown or buff brown squamules on a dirty background, involute margin, smooth or fibrillose white stipe that becomes reddish brown when bruised, negative Schäffer and KOH reactions, inferior annulus forming a broad sheath, and context that turns reddish brown when cut (Wang *et al.* 2015).

Agaricus martinicensis Pegler (1983:446) is the type species of *Agaricus* subg. *Minoriopsis. Agaricus globocystidiatus* is similar to *A. martinicensis* by the presence of cheilocystidia, by the purplish brown contents of the pileipellis, both species have the stipe covered with squamules below the annulus and a thin, membranous, pendent, white annulus that has floccose remnants of the universal veil at the lower surface. *Agaricus globocystidiatus* differs from *A. martinicensis* by the larger and beige pileus with purple scales and the presence of pleurocystidia.

The basidiomata of *Agaricus globocystidiatus* collected in Brazil share some characters attributed to *Agaricus* subg. *Minoriopsis* (Chen *et al.* 2017), including an annulus that is superous, a positive reaction to KOH, and the presence of clavate, subglobose to obovoid cheilocystidia. The other two species in the genus known to have pleurocystidia, *A. pleurocystidiatus* and *A. sinodeliciosus*, change to reddish brown upon cutting and belong to *Agaricus* subg. *Pseudochitonia* sect. *Sanguinolenti* Schaeff. & Moll. and sect. *Bivelares* (Kauffman) L.A. Parra, respectively (Heinemann 1980; Wang *et al.* 2015).

Agaricus globocystidiatus is a new species in *Agaricus* subg. *Minoriopsis* supported by morphological and molecular data. Although it is necessary to revise the morphology of the collection MATA816 (JF727870) to confirm the presence of pleurocystidia, our molecular results indicate that it is most likely *A. globocystidiatus*, which suggests a possible neotropical distribution for the new species here described. All sequences in *A. globocystidiatus* clade are 98.8–100% identical, with MATA816 presenting levels of sequence divergence between 1.2% and 0.1% compared to Brazilian sequences (MPD02 and MPD19, respectively). This work increases the knowledge of taxa within this subgenus and of *Agaricus* species that occur in America, which is important to better understand the phylogenetic relationships and the biogeographic patterns in the genus.



FIGURE 3. Agaricus globocystidiatus a-b. Basidiomata; a. FLOR61620 (MPD145); b. FLOR 61594 (MPD29-Holotype); c. Pileus surface FLOR 61599 (MPD71); d. Stipe with details of the scales and annulus with floccose lower surface FLOR61620 (MPD145); e. Pleurocystidia FLOR 61594 (MPD29-Holotype); f. Pleurocystidia (MPD04). Photos by M.P.Drewinski.

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